

Summary of thesis entitled

**PHYTOCHEMICAL STUDIES ON SOME  
CULTIVARS OF *Carica papaya* LINN. AND  
BIOPROSPECTING OF ITS RELATED SPECIES**

To

The Maharaja Sayajirao University of Baroda

For the Degree of

**DOCTOR OF PHILOSOPHY**

**IN**

**CHEMISTRY**



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### Introduction

Platelet is one of the important components of blood whose function (along with coagulation factors) is to stop bleeding by clumping and clotting the injured blood vessel [1]. Platelets, also called thrombocytes (thromb + cyte, "blood clot cell"), are anucleated, and derived from the megakaryocytes of the bone marrow. In every human being, each day about  $1 \times 10^{11}$  platelets are produced by cytoplasmic fragmentation of megakaryocytes [2]. They are 2,50,000 to 4,00,000 / $\mu$ L in human blood [2], [3] with life span of 8 to 14 days [4], [5] and are destroyed in the spleen [3]. The condition of platelet counts less than 1,50,000 / $\mu$ L is termed as 'thrombocytopenia'. The resulting symptoms are often dramatic and associated with a variety of hemorrhagic sequelae, including epistaxis, petechiae, gastrointestinal bleeding and intracranial hemorrhage.

The line of treatment for thrombocytopenia includes use of corticosteroids, immunoglobulins or splenectomy, however it is not effective for 25 to 30% of patients with chronic ITP [6], [7].

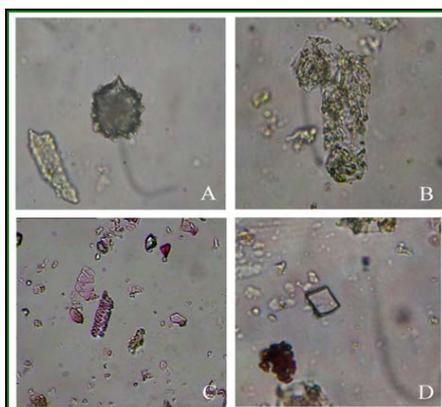
The failure of allopathic armamentarium has channelized efforts into rediscovering the wealth of traditional and complementary medicines (T&CM). While looking for effective traditional remedy for thrombocytopenia in literature, a large number of blogs describing use of *Carica papaya* for the treatment of dengue fever were seen on the social network. Dharmarathna *et al*, have described the enhancement of platelets in rats on administration of decoction of *Carica papaya* leaves [8]. *Carica papaya* leaves are traditionally used for the treatment of dengue and other ailments in Malaysia [9], [10]. Few patents on application of dry *Carica papaya* leaves as platelet booster have also been filed [11], [12].

**Inspired by these facts, an attempt was made to conduct a systematic research on an ethanopharmacologically important plant (*Carica papaya*) with a view to rectify the existing lacunae.**

To initiate any phytochemical research for development of new drug, authentication of plant material is the most crucial step. The pharmacognostic studies cover this aspect of any crude drug.

The collected plant material was identified and authenticated by Prof. M. Daniel at Department of Botany, The M. S. University of Baroda, Gujarat, India. The voucher specimen of this plant (No. BARO/2010/51) was deposited at the Herbarium, BARO, Department of Botany, The M. S. University of Baroda. Pharmacognostic studies consisting of proximate analysis, morphological studies, anatomical microscopic studies, micromorphological studies, histochemical analysis, powder studies and preliminary phytochemical analysis of the *C. papaya* leaf were performed.

The presence of sphaeraphides, starch grains and rhomboidal calcium oxalate crystals in the leaf powder form **diagnostic characters** of *C. papaya*.



**Figure 1:** Powder of *C. papaya* showing **A-** Sphaeraphide, **B-** Stomata, **C-** Spiral xylem, starch grains, **D-** Rhomboidal calcium oxalate crystal.

The preliminary screening of various groups of phytoconstituents of decoction of leaves indicated the presence of various phytochemicals like carbohydrates, proteins, amino acids, cardiac glycosides, saponins, flavonoids, alkaloids, tannins, phenolics and iridoids.

Once the authenticity of the plant was established, based on ethnopharmacological data, the crude extract of *Carica papaya* leaves were screened for its pharmacological efficacy and toxicity.

For primary screening of bioactivity, healthy Sprague Dawley rats used as targets, as *in vitro* study of platelet is not possible due to lack of nucleus. The pharmacological activity was studied by monitoring number of platelets. The results indicated increase in platelet count by about 40% irrespective of dose.

The sub-chronic activity carried out by administering 1000 mg of *Carica papaya* leaf extract per body weight of rats for 90 days revealed the *Carica papaya* leaf to be safe for oral administration.

The bioassay guided fractionation study was carried out for the polarity based chromatographically separated fractions as well as fractions separated on the basis of phytochemical groups. The results of bioassay of *Carica papaya* leaf fractions with different polarities suggested the pharmaceutically active compound to be more polar as the fractions extracted with lower polarity solvent systems did not show any significant rise in platelet counts.

The results of screening of different phytochemical groups [Table 1] indicated no conclusive rise in platelet counts with phenolics and saponin groups whereas a substantial rise in platelets is observed in case of alkaloidal extracts.

**Table 1:** Result of Screening of various phytochemical groups

| Extracts  | Platelet Count Mean±SD (10 <sup>9</sup> /L) |               |
|-----------|---|---------------|
|           | 1st day                                     | 7th day       |
| Control   | 573.33±48.23                                | 622.33±28.71  |
| Water     | 663.33±20.26                                | 791.67±13.65  |
| Saponins  | 515.33±31.26                                | 586.00±37.64  |
| Phenolics | 509.67±13.20                                | 543.00±3.61   |
| Alkaloids | 611.67±52.70                                | 1004.00±53.51 |

In order to find the lead compound responsible for the thrombocytopenic activity in *Carica papaya* leaves, thrombocytopenic wistar model was made by optimization of busulfan doses.

The various phytochemical groups were screened for the anti-thrombocytopenic activity on wistar rat model as targets and the results of screening of various phytochemical [Table 2] indicated alkaloidal group to contain the compound/s we were looking for.

**Table 2:** Result of screening of phytochemical group on antithrombocytopenic model

| Extracts  | Platelet Count Mean±SD (10 <sup>9</sup> /L) |               |
|-----------|---|---------------|
|           | 1st day                                     | 20th day      |
| Control   | 623.25±105.32                               | 704.75±132.92 |
| Diseased  | 800.75±142.57                               | 78.75±27.28   |
| Water     | 782.50±189.19                               | 33.50±15.24   |
| Saponins  | 883.75±133.30                               | 112.50±24.12  |
| Phenolics | 813.50±153.21                               | 72.25±17.69   |
| Alkaloids | 973.00±243.96                               | 313.25±50.74  |

On the basis of these observations, the alkaloidal fraction was further sub-fractionated and the anti-thrombocytopenic activity on the model animal target was studied. The bioassay guided fractionation study revealed that the active pharmacological component present in the *Carica papaya* leaves with anti-thrombocytopenic activity belongs to the alkaloidal group and it could be carpaine, the chief alkaloid component [Table 3].

**Table 3:** Results of screening of sub-fractions of alkaloidal group on antithrombocytopenic model

| Extracts                            | Platelet Count Mean±SD (10 <sup>9</sup> /L) |               |
|-------------------------------------|---|---------------|
|                                     | 1st day                                     | 20th day      |
| Control                             | 748.50±203.96                               | 718.75±157.89 |
| Diseased                            | 730.25±135.53                               | 81.50±30.21   |
| Pet. Ether fraction of alkaloids    | 660.50±99.47                                | 759.75±181.44 |
| Ethyl acetate fraction of alkaloids | 578.25±54.79                                | 632.25±116.23 |
| Carpaine (Single compound)          | 623.00±168.74                               | 555.50±42.99  |

To understand the apparent better activity of the petroleum ether fraction, quantitation of carpaine in the sub-fractions was thought to be most appropriate approach. To serve the afore said purpose, an LCMS–MS method was developed and validated.

The analysis revealed quantity of carpaine in water, petroleum ether and ethyl acetate extract to be  $6.46 \pm 0.09$  mg/g,  $28.56 \pm 0.59$  mg/g,  $25.08 \pm 0.53$  mg/g respectively.

Once the lead pharmacological group was identified, the next study was about bioprospecting of carpaine in different parts of the plant and **Chapter 4** is about isolation, characterization and quantification of carpaine in different parts of *Carica papaya*.

A modified method by optimizing various parameters i.e. percentage of acetic acid, pH, temperature, bases and time was developed for the extraction of carpaine from different parts of plant. Due to non-chromophoric nature of carpaine it was very difficult to analyze using HPLC with PDA or UV detector, Hence new LCMS method developed for quantification of carpaine was found to be fast, simple and sensitive with good precision and accuracy. The figure of merit [Table 4] for the method were as follows

**Table 4:** Regression data, LOD, LOQ for carpaine

| m/z | component | Regression    | r <sup>2</sup> | LOD(ng/mL) | LOQ(ng/mL) |
|-----|-----------|---------------|----------------|------------|------------|
| 479 | Carpaine  | y=22700x-23.8 | 0.9991         | 0.09       | 0.25       |

The intraday (n=5) and inter day (n=9) analysis precision RSD% for intraday was 1.35 and precision RSD% for interday was 2.09. The accuracy (recovery) (n=3) was 89.1.

The LCMS-MS analysis confirmed that all the parts of papaya plant contained carpaine. The results as shown in table 5 indicated leaf to contain highest concentration of carpaine followed by fruit pulp, fruit peel, roots, stem, petioles and seeds. Results also shows there was no significant difference between the carpaine content among different age group of leaves.

**Table 5:** Quantity of carpaine present in different parts of papaya plant

| Papaya parts           | Content (mg/g) |
|------------------------|----------------|
| Leaves (mature)        | 9.30           |
| Fruit pulp             | 4.90           |
| Fruit Peel             | 1.99           |
| Roots                  | 1.86           |
| Stem                   | 1.04           |
| Petioles               | 0.72           |
| Seeds                  | 0.65           |
| Tendril leaves         | 0.76           |
| 2-3 weeks older leaves | 0.57           |

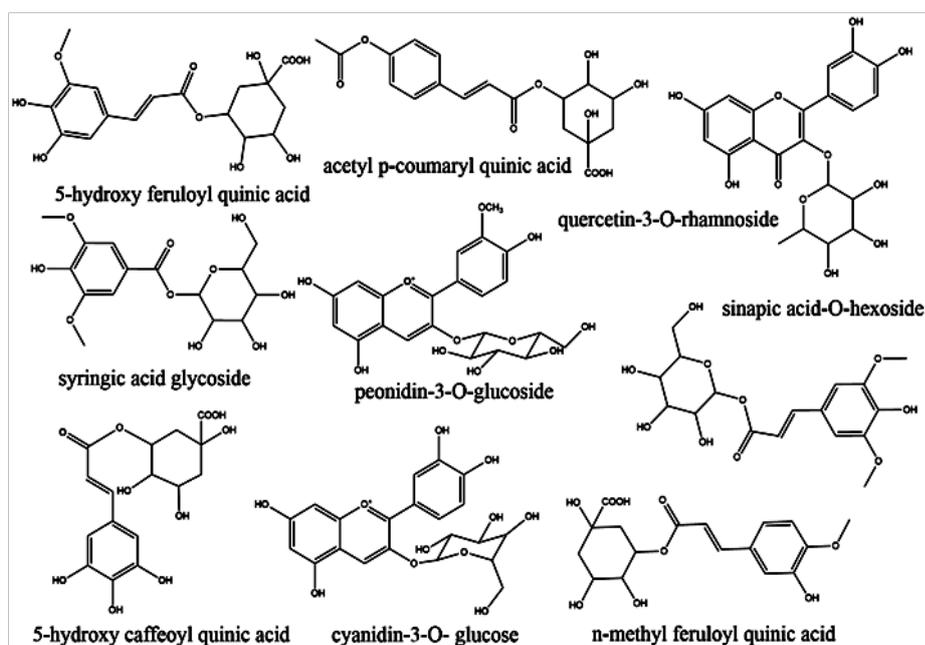
*Carica papaya* has abundant phenolics in different parts. There are few reports on the study of antioxidant activities of phenolics in different cultivars of *Carica papaya* [13]–[20].

However the study of comparative study of phenolics in different parts of *Carica papaya* has not been reported, hence isolation and identification of phenolics different chromatographic techniques were used including 1D and 2D paper chromatography, HPLC and LCMS-MS method were carried out [Chapter 5].

The highly polar phenolic groups that gave bluish-white fluorescence with  $R_f \sim 1$  were directly used for qualitative analysis [21]. The 2-D paper chromatography indicated presence of different phenolics (mainly protocatechuic acid) with different colour forming derivatives with p-nitro aniline and sulphanic acid.

Bioprospecting of phenolics among different parts using HPLC confirmed the presence of protocatechuic acid, chlorogenic acid, caffeic acid, p-coumaric acid, syringic acid, p- hydroxy benzoic acid.

The LCMS-MS analysis of isolated phenolic fractions of aqueous extract indicated presence of twenty compounds belonging to a typical hydroxy cinnamic acid and phenolic acid derivatives.



. **Figure 2:** New Phenolics identified from different parts of papaya plant

The quantitative analysis of phenolics and flavonoids along with their antioxidant activities of different parts of papaya plant were performed by total flavonoid content (TFC) and total phenol content (TPC).

**Table 6:** DPPH scavenging activity, FRAP activity, total flavonoid content (TFC) and total phenol content (TPC) of different parts of *C. papaya* plant

| <b>Samples</b> | <b>DPPH<br/>IC<sub>50</sub>(mg/mL)</b> | <b>FRAP<br/>IC<sub>50</sub>(mg/mL)</b> | <b>TPC ± SE<br/>(µg GAE/mg)</b> | <b>TFC ± SE<br/>(µg Qtn/mg)</b> |
|----------------|--|--|---------------------------------|---------------------------------|
| <b>Seed</b>    | 3.11 ± 0.11                            | 4.85 ± 0.27                            | 865.73 ± 0.35                   | 31.41 ± 0.01                    |
| <b>Fruit</b>   | 3.36 ± 0.13                            | 12.43 ± 0.34                           | 981.53 ± 0.53                   | 22.99 ± 0.02                    |
| <b>Leaf</b>    | 13.31 ± 0.48                           | 13.54 ± 0.71                           | 815.29 ± 0.48                   | 9.09 ± 0.01                     |
| <b>Root</b>    | 13.87 ± 0.08                           | 48.15 ± 2.59                           | 315.59 ± 0.19                   | 3.48 ± 0.01                     |
| <b>Stem</b>    | 39.48 ± 0.4                            | 94.24 ± 9.48                           | 298.34 ± 0.11                   | 5.82 ± 0.01                     |

Thus the thesis that *Carica papaya* leaves contains some component that is responsible for the antithrombocytopenic activity has been proved through systematic bioassay guided fractionation and the phytochemical responsible for pharmacological activity is alkaloid (Carpaine), which is present in all parts of the plant and is maximum in the *Carica papaya* leaf.

### References

- [1] K. Laki, "Our ancient heritage in blood clotting and some of its consequences," *Ann. N. Y. Acad. Sci.*, vol. 202, no. 1, pp. 297–307, 1972.
- [2] O. Garraud and F. Cognasse, "Are platelets cells? And if yes, are they immune cells?," *Front. Immunol.*, vol. 6, no. 70, pp. 1–8, 2015.
- [3] M. Gawaz, *Blood Platelets: Physiology, Pathophysiology, Membrane Receptors, Antiplatelet Drugs, Coronary Heart Disease, Stroke, Peripheral Arterial Disease ; 47 Tables*. Thieme, 2001.
- [4] G. O. Evans, *Animal Hematotoxicology: A Practical Guide for Toxicologists and Biomedical Researchers*. CRC Press, 2008.
- [5] Y. Najean, N. Ardaillou, and C. Dresch, "Platelet lifespan.," *Annu. Rev. Med.*, vol. 20, pp. 47–62, 1969.
- [6] C. Neunert, W. Lim, M. Crowther, A. Cohen, L. Solberg, and M. A. Crowther, "The American Society of Hematology 2011 evidence-based practice guideline for immune thrombocytopenia," *Blood*, vol. 117, no. 16, pp. 4190–4207, 2011.
- [7] S. L. Spitalnik, S. Arinsburg, and J. Jhang, *Clinical Pathology Board Review*. Elsevier Health Sciences, 2014.
- [8] S. L. C. A. Dharmarathna, S. Wickramasinghe, R. N. Waduge, R. P. V. J. Rajapakse, and S. A. M. Kularatne, "Does *Carica papaya* leaf-extract increase the platelet count? An experimental study in a murine model," *Asian Pac. J. Trop. Biomed.*, vol. 3, no. 9, pp. 720–724, Sep. 2013.
- [9] K. Sathasivam, S. Ramanathan, S. M. Mansor, M. R. M. H. Haris, and W. H. Wernsdorfer, "Thrombocyte counts in mice after the administration of papaya leaf suspension," *Wien. Klin. Wochenschr.*, vol. 121, no. 3, pp. 19–22, 2009.
- [10] S. Subenthiran, T. C. Choon, K. C. Cheong, R. Thayan, M. B. Teck, P. K. Muniandy, A. Afzan, N. R. Abdullah, and Z. Ismail, "Carica papaya Leaves Juice Significantly Accelerates the Rate of Increase in Platelet Count among Patients with Dengue Fever and Dengue Haemorrhagic Fever," *Evidence-based Complement. Altern. Med.*, vol. 2013, pp. 1–7, 2013.
- [11] J. Alva and M. Thapar, "Increasing low platelets instantly," WO/2010/041263, 2010.

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- [12] A. B. M. F. Karim, C. M. Hasan, H. Sohrab, M. A. Al-Mansur, and R. B. Karim, "Product and method for treating thrombocytopenia," WO/2011/028098, 2011.
- [13] G. Srikanth, S. M. Babu, C. H. N. Kavitha, M. E. B. Rao, N. Vijaykumar, and C. H. Pradeep, "Studies on in-vitro antioxidant activities of Carica papaya aqueous leaf extract," *Res. J. Pharm. , Biol. Chem. Sci.*, vol. 1, no. 2, pp. 59–65, 2010.
- [14] A. Özkan, H. Gübbük, E. Güneş, and A. Erdoğan, "Antioxidant capacity of juice from different papaya ( Carica papaya L .) cultivars grown under greenhouse conditions in Turkey," *Turkey J. Biol.*, vol. 35, pp. 619–625, 2011.
- [15] K. Zhou, H. Wang, W. Mei, X. Li, Y. Luo, and H. Dai, "Antioxidant activity of papaya seed extracts," *Molecules*, vol. 16, no. 8, pp. 6179–6192, 2011.
- [16] F. O. Asmah R, "Proximate Analysis, Antioxidant and Anti Proliferative Activities of Different Parts of Carica papaya," *J. Tissue Sci. Eng.*, vol. 5, no. 1, pp. 1–7, 2014.
- [17] Z. Tahir, M. Arshad, and S. K. Chaudhari, "Redox protective potential of fruits and vegetables: A review," *J. Coast. Life Med.*, vol. 3, no. 8, pp. 663–668, 2015.
- [18] A. Maisarah, B. Nurul Amira, R. Asmah, and O. Fauziah, "Antioxidant analysis of different parts of Carica papaya.," *Int. Food Res. J.*, vol. 20, no. 3, pp. 1043–1048, 2013.
- [19] N. Asghar, S. A. R. Naqvi, Z. Hussain, N. Rasool, Z. A. Khan, S. A. Shahzad, T. A. Sherazi, M. R. S. A. Janjua, S. A. Nagra, M. Zia-Ul-Haq, and H. Z. Jaafar, "Compositional difference in antioxidant and antibacterial activity of all parts of the Carica papaya using different solvents.," *Chem. Cent. J.*, vol. 10, p. 5, 2016.
- [20] Z. Radhi Addai, A. Abdullah, and S. Abd Mutalib, "Effect of extraction solvents on the phenolic content and antioxidant properties of two papaya cultivars," *J. Med. Plants Res.*, vol. 7, no. 47, pp. 3354–3359, 2013.
- [21] A. J. Harborne, *Phytochemical Methods A Guide to Modern Techniques of Plant Analysis*. Springer Netherlands, 1998.
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