
CHAPTER-1

GENERAL INTRODUCTION

CHEMISTRY OF GENUS ASPARAGUS

In the systematic classification, genus Asparagus belongs to the family Liliaceae and its position in the plant kingdom¹ is shown below.

Division	...	Trachaeophyta
Sub-division	...	Pteridopsida
Class	...	Angiospermae
Sub-class	...	Monocotyledoneae
Order	...	Liliales
Family	...	Liliaceae
Genus	...	Asparagus

The name Asparagus¹⁻⁸ was applied by Theophratus. The genus Asparagus comprises of erect climbing herbs or shrubs with slender, ribbed or grooved green stems. These have no true leaves, but small, sometimes spine-tipped, scales subtending a bunch of narrow green needle-like spiny or flattened branchlets. (cladodes). Plants of genus Asparagus are found in India, Ceylon, Burma, Java, Africa and Europe.

The species coming under the genus Asparagus are of medicinal value. The leaves and the roots are usually used for a number of diseases. They contain carbohydrates, saponin, saponins, proteins, vitamins, amino acids, lipids and inorganic materials like Mg, Fe, K, Na P, P₂O₅, K₂O etc.

The genus *Asparagus* covers over a hundred species. The following species are more frequently met with.

- (1) *A. racemosus* Willd.
- (2) *A. officinalis* Linn.
- (3) *A. asparagoides* Wight.
- (4) *A. sprengeri* Regel.
- (5) *A. filicinus* Buch-Ham.
- (6) *A. acutifolius* Linn.
- (7) *A. adscendens* Roxb.
- (8) *A. gonoclados* Baker.
- (9) *A. umbellatus* Linn.
- (10) *A. stipularis* Forsk.
- (11) *A. curillus* Ham.
- (12) *A. cochinchinensis* (Loureio) Merrill.

1. *A. racemosus* Willd

This is a tall spinuous climbing, excessively branched under-shrub with small white flowers which are very fragrant. The roots are tuberous. This species is found in tropical and subtropical India, Ceylon, tropical Africa, Java and Australia.

The root of this plant is used medicinally as a refrigerant, demulcent, diuretic, aphrodisiac, antispasmodic, alterative, antidiarrhoeatic and antidysentric. It is used chiefly as a demulcent in veterinary medicine.

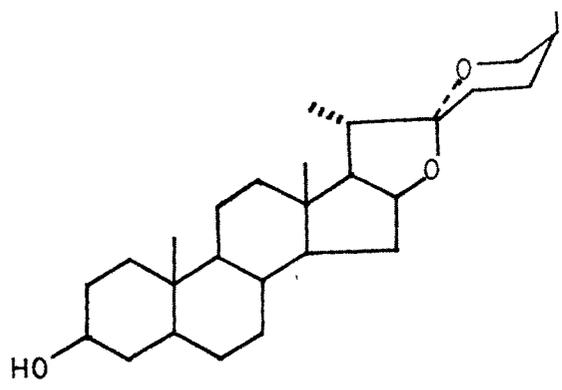
It prevents confluence of small-pox. It is considered to be a galactogogue. The native physicians used the root as a stimulant and restorative.

Rao⁹ et al. reported that the root powder has the following composition.

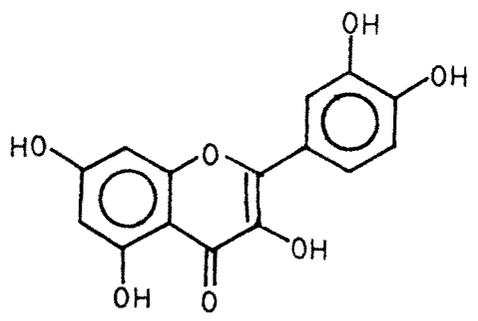
Moisture	...	13.42%
Lipids	...	3.19%
Proteins	...	2.95%
Saponins	...	5.44%
Carbohydrates) including) uronic acids) and free sugars)	...	52.89%
Crude fibre	...	17.93%
Ash	...	3.92%

The quantitative analysis of the ash revealed the presence of Na, K, Ca, Mg, Fe, CO₃^{''}, SO₄^{''}, PO^{'''} and silica.

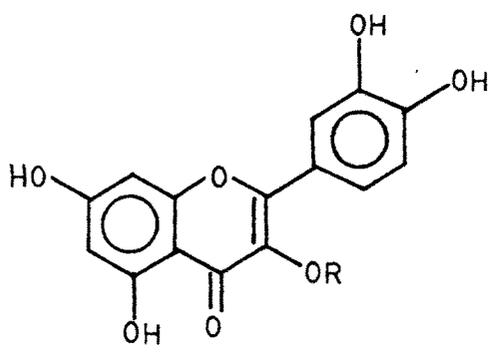
The root powder freed from saponins by treatment with ethylacetate was extracted with 70% alcohol. The extract contained both mono and oligosaccharides (37.44%). The sugars obtained after hydrolysis of the extract were identified as glucose and fructose. The mucilage⁹⁻¹¹ isolated after the removal of saponins and free sugars was composed of glucose and galactose in the proportion of 3:2.



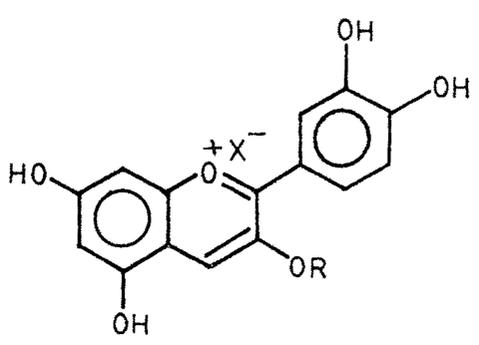
I



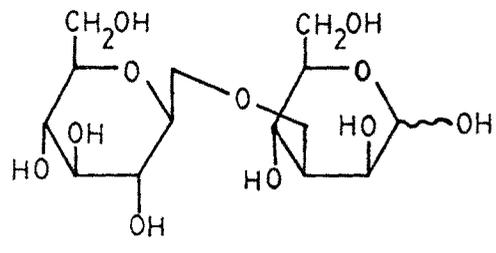
II



III $R = C_{12}H_{21}O_9$
= 6-(α -L-rhamnosyl)-D-glucoside



IV $R = \text{galactosyl group } (C_6H_{11}O_5)$
 $R_2 = \text{glucosyl rhamnoside } (C_{12}H_{21}O_9)$



VI

FIG. I

Rao¹² isolated sarsasapogenin (I) from the acid hydrolysate of the alcohol extract of defatted root powder.

Subramanian¹³ et al. isolated quercetin (II), rutin (III) and hyperoside from the flowers. No quercetin was found in the fruits. In fully ripe fruits cyanidin-3-galactoside (IV) and cyanidin-3-glucorhamnoside (V) were shown to be present.

From the ethanolic extract D-glucose, D-mannose and a disaccharide (VI) m.p. 164-5°, were isolated¹⁴. Subramanian¹⁵ et al. obtained diosgenin (VII), m.p. 204-5° and quercetin-3-glucuronide from leaves.

2. A. officinalis Linn.

This is the commonly known Asparagus. This species is usually erect, much branched herb with yellowish green flowers. The berries are red and the roots fleshy.

Berries are used⁶ by Hakims in debility of the stomach and also in liver, spleen and renal disorders. It is a tonic and an aphrodisiac. The leaves, berries and the roots are used as demulcent and diuretic. It is also used in cardiac dropsy and chronic gout¹⁶. Also the branched lower stems are excessively eaten as a vegetable all over

the world. The plant¹⁷ is diuretic, laxative, cardiac and sedative. The infusion made from the roots will help against jaundice and congestive torpor of the liver.

In England, a medicinal tincture is made from the whole plant which allays minory irritation and does good against rheumatic gout. A syrup of asparagus is employed medicinally in France. The water in which asparagus has been boiled is beneficial against rheumatism. In U.S.A. asparagus is thought to be undeniably sedative and a palliative in all heart affections attended with excited action of the pulse.

Asparagus officinalis is one of the most widely studied species. It is rich in vitamins. It contains¹⁶:

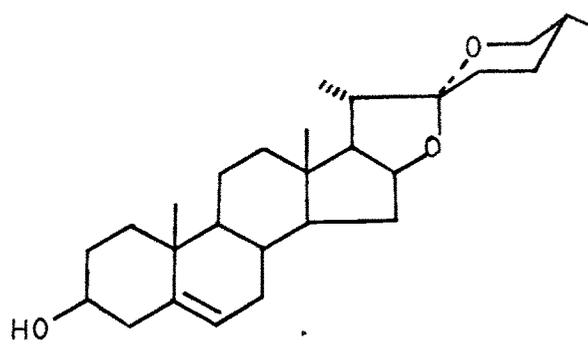
Moisture	...	93%
Protein	...	2.2%
Fat	...	0.2%
Carbohydrates	...	3.2%
Fibre	...	0.7%
Ash	...	0.7%
Ca	...	0.025%
P	...	0.039%

Fe, 0.96 mg, Cu, 0.14 mg.

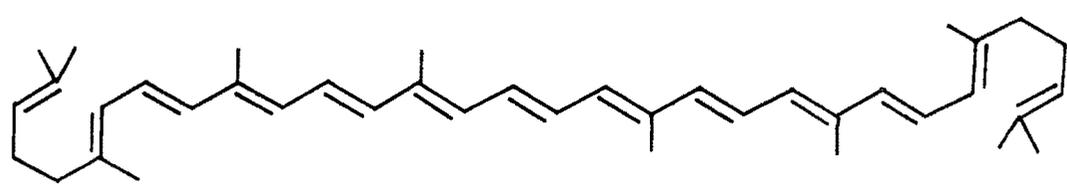
Vitamins - A, 1400 I.U.; B₁, 180 g.; B₂, 130 g.; C, 40 mg/100 g. (Heinz Co. Nutrit. Charts. 1942, 21).

Tollens¹⁸ reported the following composition from the analysis of the plant.

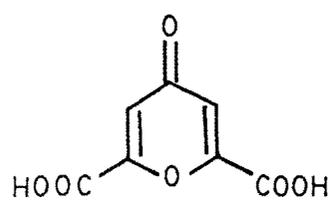
V7



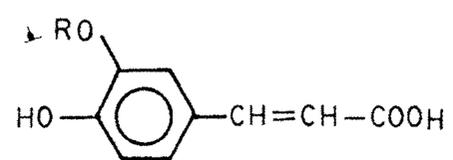
VII



VIII

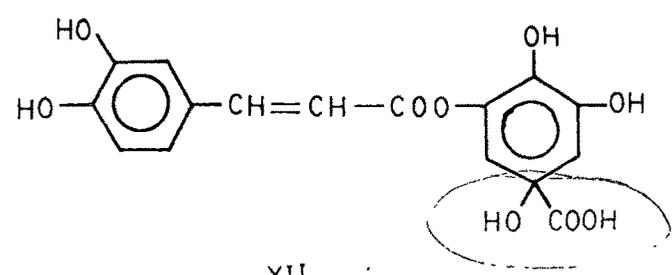


IX



X R= CH₃

XI R= H



XII

Water	...	4.4%
Fat	...	1.5%
Fibre	...	12.4%
Sugars	...	31.5%
Pentosans	...	8.6%
P ₂ O ₅	...	1.0%
K ₂ O	...	3.0%

Tollens¹⁹ further showed that roots contained fructose and glucose. An oil containing mainly palmitic acid was isolated²⁰ from the root. The mucilage isolated from the roots was composed of glucuronic acid, glucose, fructose and mannose in the ratio of 1:8:2:10²¹.

A white crystalline powder, Asparagin²² C₄H₈O₃N₂·H₂O was found to be present in all parts of the plant. Goldberg²³ isolated a hemicellulose, made of β-glucose and β-mannose (1→4) with α-galactose as the terminal group.

From the fruit, Hehner²⁴ obtained carbohydrates, organic acids such as maleic and citric acids, and an ash which on partial analysis, showed SiO₂, Al₂O₃, Fe₂O₃, CaO, Mg, K₂O, Na₂O, P₂O₅. Bartlett²⁵ et al. reported the presence of glucose, fructose, galactose and mannose in

the hydrolysate of hemicellulose, isolated from the seeds. Lycopine (VIII) was also reported²⁶ in the berries.

Tanret²⁷ isolated two carbohydrates (Asparagose, $(C_6H_{10}O_5)_n$, m.p. 198-200°; pseudo-asparagose - both composed of glucose and levulose). Mannitol was reported²⁸ to be present in the juice of the plant.

Chelidonic acid²⁹ (IX) and β, β' -dimercaptoisobutyric acid³⁰, ferulic acid³¹ (X), caffeic acid³¹ (XI) and chlorogenic acid³¹ (XII) were also reported to be present.

Flavonoids³² such as quercetin (II), rutin (III) anthocyanins³³, like cyanidin-3-rhamnoglucosyl glucoside (XIII), cyanidin-3-rhamnosyl glucoside (XIV), peonidin-3-glucosyl rhamnosyl glucoside (XV) and peonidin-3-rhamnosyl glucoside (XVI) were isolated.

Eight oligosaccharides³⁴ were isolated from purified oligosaccharide fractions of the roots. The roots contained 9 steroid glycosides³⁵ (TLC, column and paper chromatography) which are named asparagosides A, B, C, D, E, F, G, H and I. From methanol extract of the roots -sitosterol, sarsasapogenin and 9 glycosides were isolated³⁶. Asparagosome A, C, D and F contained spirostanol³⁷ as their aglycone, whereas asparagosides B, E, G, H and I contained (25 S)-5 β -furostan-3 β , 22 α ,

26-triol^{37,38} as their aglycone. Asparagosides A, E and G contained glucose as their only sugar whereas asparagosides F, H and I had xylose alongwith glucose.

The species also contained glycoside of 22-spirostan-3 β -ol, rhamnose, xylose and glucose and 22-isospirostan-3 β -ol³⁹.

From the edible part Held⁴⁰ et al. isolated diosgenin (VII) and yamogenin (XVII).

The other components reported to be present in the species are the following: inorganic matter⁴¹, such as Na, K, Ca, Fe, Mg, Cu, P, ascorbic acid⁴², thiamine⁴², riboflavin⁴², Carotene⁴², folic acid⁴¹, choline⁴¹, nicotinic acid⁴¹, pantothenic acid⁴¹, inositol⁴¹, fumaric acid⁴³, succinic acid⁴³, α -ketoglutaric acid⁴³, pyrrolidone carboxylic acid⁴³, glycolic acid⁴³, maleic acid and citric acid⁴³ and amino acids⁴⁴ such as valine, leucine, isoleucine, threonine, arginine, histidine, lysine, phenyl alanine, tryptophan and methionine.

A water soluble⁴⁵ galactoglucomannan was isolated from endosperm of A. officinalis. It contains residues of glucose, mannose and galactose, in the ratio of 43:49:7.

3. A. asparagoides Wight.

This species is a tall, slender, glabrous twiner. It is much branched. The flowers are greenish white and the berries dark purple. These are grown much for florists' use in decoration.

Diosgenin (VII) and pennogenin (XVIII) were isolated⁴⁶ from the rhizomes of this species.

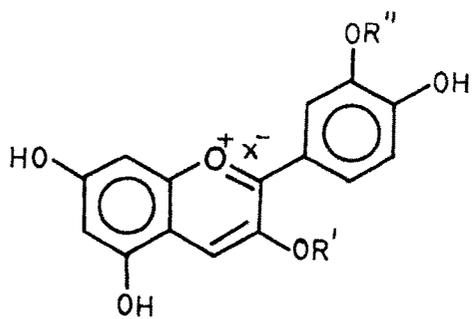
4. A. sprengeri Regel.

This is a climbing and much branched species with small prickles. The flowers are small white pink and fragrant. Berries are bright coral red. It is a good room plant. The roots are tuberous.

Elze⁴⁷ had isolated an oil from the flowers by solvent extraction. The oil had strong aldehydic odour and yielded a semicarbazone. Held⁴⁰ et al. isolated diosgenin (VII) and yamogenin (XVII) from the hydrolysate of the methanol extract.

5. A. filicinus Buch-Ham.

This species is a tall, erect herb. It is highly branched, smooth unarmed with the lower branches spreading.

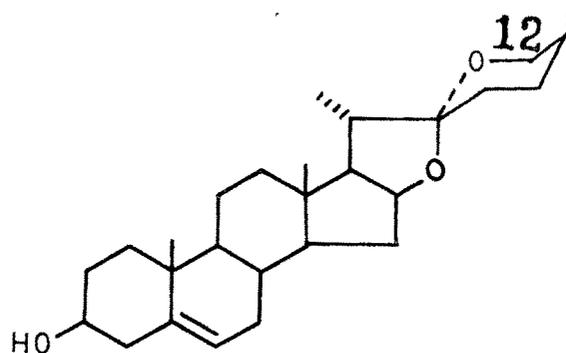


XIII $R' =$ rhamno glucosyl glucoside group
 $R'' = H$

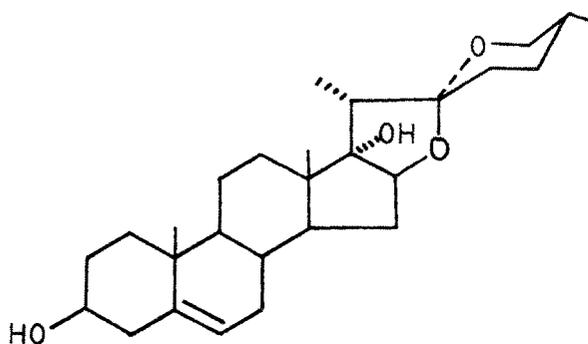
XIV $R' =$ rhamnosyl glucoside group
 $R'' = H$

XV $R' =$ glucosyl rhamnosyl glucoside group
 $R'' = CH_3$

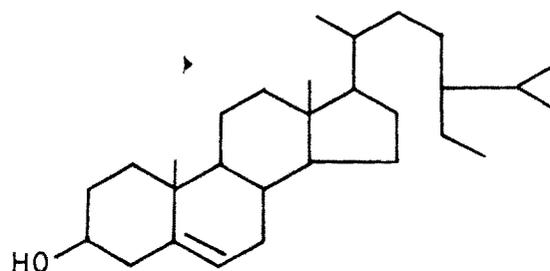
XVI $R' =$ rhamnosyl glucoside group
 $R'' = CH_3$



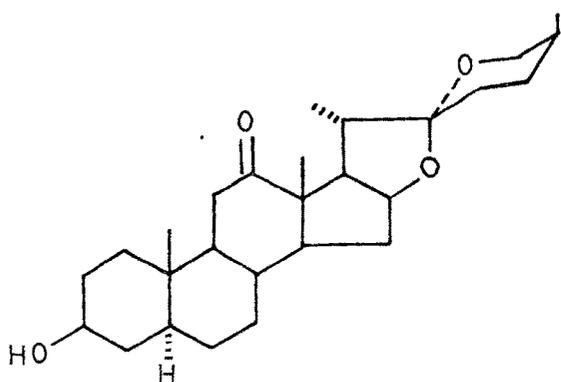
XVII



XVIII



XIX



XX

The flowers are green and the berries black. Roots are tuberous and are densely clustered. It is found in Kashmir, Bhutan, The Khasia Hills, Assam, Burma and China.

The root is considered⁶ as a tonic and astringent. A spring of this plant is put in the hands of small-pox patients as a curative measure. It is aphrodisiac. The root⁴⁸ is considered vermifuge and taeniafuge in Annam. It is given in cholera and acts as a powerful diuretic. It is also used as a cure for rheumatism due to dampness.

Rao^{49,50} et al. isolated an unidentified colouring matter, glucose, fructose and a mucilage composed of mannose, fructose, glucose (5:4:1) and a uronic acid (mannoglucuronic acid) from the powder.

6. A. Acuitifolius Linn.

It is a woody shrub with zigzag branching and yellow flowers. The berries are waxy olive-green. It is found in Mediterranean region.

From roots and rhizomes, sarsasapogenin (I) has been isolated⁵¹⁻⁵³.

7. A. adscendens Roxb.

The species is a tall, stout, excessively branched shrub. The branchlets are grooved ascending and angled. The flowers are white. It is found in Western Himalaya, Punjab, Murre, the Dieu and Sal forests.

It is sometimes used as a vegetable¹⁶. The white tubers are hairy and mucilagenous and swell up with water. They are reported to possess cooling and demulcent properties. They are used as tonic and also as a substituent for Salep. They are useful in diarrhoea, dysentery and general debility.

The powdered roots contain essential oil which smells like Khus oil. Rao⁵⁴ et al. analysed the tuber powder and found the following.

Moisture	...	11.40%
Fat	...	0.87%
Saponins	...	5.02%
Proteins	...	5.44%
Carbohydrates including free sugars and uronic acids)))) ...	46.84%
Crude fibre	...	23.42%
Ash	...	6.28%

Analysis of the ash showed the presence of K, Na, Fe, CO₃["], SO₄["], PO₄["] and silica.

The mucilage isolated, constituted of glucose, mannose, xylose and glucuronic acid.

Two hemicelluloses were isolated by Rao⁵⁵ et al. one of which was named as A₂, was shown to contain xylose, glucose and glucuronic acid in the ratio of 2:1:1 while the other one, B₂ was shown to have the same sugars in the ratio of 1:1:2.

Sitosterol, sarsasapogenin and diosgenin have been reported⁵⁶ from the fruits of this plant. Two furostanol and two spirostanol glycosides are reported from the methanol extract of the fruits⁵⁶ on acid hydrolysis they gave sarsasapogenin. Three of them had glucose as the only sugar, whereas one contained glucose and rhamnose.

The methanol extract of the defatted roots of this plant yielded⁵⁷ β -sitosterol- β -D-glucoside, two new spirostanol glycosides named as asparanin C and asparanin D, and two new furostanol glycosides asparoside C and asparoside D.

8. A. gonoclados Baker.

This is a much branched undershrub. Flowers are white. The green branches are curved and strongly angled. It is found in Konkan, Kanara, Western Ghats of Madras State, Ceylon etc.

The root is considered as nourishing and aphrodisiac. Boiled with oil, it is applied to cutaneous diseases. It is given in gonorrhoea in 15 grains per dose. The root is used as an adulterant or as a substitute for Aconitum heterophyllum.

The plant contains saponins. Sarsasapogenin (I) was isolated from the acid hydrolysis of the alcohol extract¹².

9. A. umbellus Link.

This is a spreading climber or undershrub with branches and twigs reflexed and pendulous. The flowers are white and the berries yellow to dark red. The plant is used by florists for decorative purpose.

From the rhizomes, A. Gonzalez⁵⁸ et al. isolated β -sitosterol (m.p. 134-8°, XIX), tigogenin (m.p. 213-4°), hecogenin (m.p. 267°, XX) and hispidogenin.

10. A. stipularis Forsk.

This species is a climbing and much branched one. The branches are angled and grooved with prickles. The flowers are white. It is abundantly found in Mediterranean region.

Panizo⁵³ et al. isolated sarsasapogenin (I) from rhizomes.

11. A. curillus Ham.

This is an excessively branched undershrub. It is widely occurring in subtropical Himalaya from Kumaon to Nepal region.

Sharma⁵⁹ and his coworkers reported two spirostanol and two furostanol glycosides from the fruits of this plant. From methanol extract of the roots, Sharma⁶⁰ et al. isolated three spirostanol and two furostanol glycosides.

12. A. cochinchinensis (Loureiro) Merrill⁶¹

The dried fruits of this plant are known as Asparagi radix. The plant occurs widely in Japan and China. Tuberos roots of this plant have been used in Chinese crude drug for the purpose of analeptic, diuretic and cough medicine. It is also used as a tonic and exhibits antifebrile, antitussive and diuretic activities.

β -Sitosterol, glucose, and fructose have been reported from this plant⁶². From the roots, eight oligosaccharides⁶³ constituted of glucose and fructose have been isolated.

Okanishi Tameo⁶⁴ et al. reported sarsasapogenin (I) from roots of the plant. Four Ehrlich - positive⁶⁵ furostanol oligosides have been reported by Tenji Konishi and Junzo Shoji⁶⁶.

Tomoda⁶⁷ et al. isolated citrulline, asparagin, serine, threonine, proline, glycine, alanine, valine, methionine, leucine, isoleucine, phenyl alanine, tyrosine, asparatic acid, glutamic acid, arginine, histidine, lysine and an unidentified acidic amino acid.

SHATAVRI IN AYURVEDA

The roots of shatavri are used as a medicine in Ayurvedic system for last several centuries, as is described in Charaka Samhita, Sushrut Samhita and other books of Ayurveda.

In Ayurvedic system shatavri is a well known drug having a number of properties. In ayurvedic literature it is referred to as by different synonames, the names are characteristic and indicative of its properties. In Ayurveda Saukhyam of Todarananda⁶⁸ it is described as satavari, bhirupatri, dvipika, adharkantaki, narayani, satapadi, satahva and vahuputrika. The synonames used are shatamooli, sheetvirya, bahusutaa, atirasa, abhiru,

maha purusa, dantika, sahastra virya, kesi, tungini and sukmapatrika. In ayurved literature its actions are described⁶⁹ as balya (promoter), vedanasthapana, rasayana (rejuvenation) and stanyakari.

Shatavari is either used singly⁷⁰⁻⁷² or as an ingredient in the kalp⁷⁰⁻⁷². The root is highly mucilagenous, anti-diarrhoeatic, refrigerant, diuretic, antidysentric, nutritive tonic, demulcent, galactogogue, aphrodisiac and anti-spasmodic⁷³. In addition shatavari is laxative, and expectorant⁷⁴. It is given⁷⁵ in billiousness, rheumatism, dyspepsia, in combination with other diuretics, it is given in scanty urine. As a tonic it is given in sexual debility and respiratory complaints. It is used⁷⁶ as a cardiac tonic and it increases intellectual power, also used in chronic colitis and hemorrhoids. Sushruta recommends the use of shatavari in internal piles. Arya Bhishaka, Charakadatta, Bhavaprakash recommended it in hyperacidity, bilinary colic, burning syndrom and hemorrhagic diseases. Bhavaprakash and Charaka recommended it as nervine sedative and in epilepsy. Charakadatta has recommended it for gout. It is useful in urinary calculus, in the diseases of blood and the eye, throat complaints, tuberculosis, leprosy and night blindness. It is used as an ingredient in a mixture which cures rakta-pitta, anorexia, suppression of the power of digetion,

vomitting, anemia, jaundice, syphilis, erysipelas and chronic fever⁷⁷. It is also used in scalding urine, gleet and gonorrhoea. The root of this plant is used as a demulcent in veterinary medicine. Combined with other ayurvedic drugs, it cures oedema, chronic tuberculosis; immobility of arm and facial paralysis, asthma, hiccup, cough, insanity, syncope, ascitis, chlorosis and constipation. Also it is antiabortive and increases virility and promotes fertility.

BOTANICAL IDENTIFICATION AND PHARMACOGNOSTIC STUDIES

The drug has been equated with various species of Asparagus by many authors. However, A. racemosus Willd. of the family of liliaceae resembled most with shatavari. The taxonomical characters prescribed for A. racemosus are the characters of spines, size of clusters and the length of the cladodes.

78-79

Kanitkar et al. made a detailed pharmacognostic and microscopic study of the authentic A. racemosus Willd. which were obtained from different parts. They found that there are a number of discrepancies in respect of character of spines, size of clusters and the length of the cladodes. According to their study, the classification based on the characters like length of cladodes, size of cladode clusters and size of spines are subject to change according to their

position on the plant and growth of the plant in different soils and climate is inadequate for comparison. They suggested pith/stele ratio as a distinct character of shatavari materials.

PHARMACOLOGICAL SCREENING OF DIFFERENT EXTRACTS OF
SHATAVARI

Various extracts of shatavari were prepared by extracting the material with ethyl alcohol, ethyl acetate, petroleum ether and acetone. The alcohol extract was found to contain saponins and free sugars. The extraction procedure was modified. The roots were first extracted with hot acetone and then with hot alcohol in a soxhlet unit. The alcohol extract, after removal of solvent, was dissolved in water and extracted with n-Butyl alcohol saturated with water. The butanol extract contained glycosides - i.e. saponin fraction.

Gaitonde⁸⁰⁻⁸³ et al. carried out the pharmacological screening of various extracts and found that the alcohol extract had the antioxytocio activity.

TABLE - 1 PHARMACOLOGICAL SCREENING OF VARIOUS EXTRACTS OF SHATAVARI⁸⁴

Extract	Diuretic	Anabolic	Anti-ADH activity	Antioxy - toxic activity	Mamma - rotropic activity	Human toxicity study
Alcohol	-	-	-	Present	Present	-
Ethylacetate	-	-	-	Present	Present	-
Acetone	-	-	-	Present	Present	-
Petroleum ether	-	-	-	Absent	Present	-
Saponin mixture	-	Absent	-	Present	Present	-
Saponin A ₄	-	-	Present	Present	Absent	No
(Shatavarin I)						Toxicity

Roy⁸⁵ et al. reported that the crude alcohol extract of A. racemosus caused an initial increase in force and rate of contraction in isolated frog's heart but a higher dose of crude alcohol extract produced a fall in the blood pressure and depression in the respiration of cat. The extract also caused an increase in bleeding time in the rabbit and slight diuretic effect in rat⁸⁶.

The alcohol extract of aerial parts of A. racemosus showed anti-cancer activity⁸⁷.

The aqueous extract of both fresh and dried roots were found to have amylase and lypase activities⁸⁸.

R E F E R E N C E S

1. E.L. Core, Plant Taxonomy p. 215, Prentice-Hall Inc., New Jersey (1962).
2. J.D. Hooker, Flora of British India Vol.VI, p. 314, First Indian Reprint (1973).
3. F.J. Chittenden, Dictionary of Gardening Vol.I, p. 193 Oxford, Clarendon Press, 1951.
4. P.Y. Fysen, Flora of South Indian Hill Stations Vol.I, p. 608, Government Press, Madras.
5. R. Hegnauer, Chemotaxonomy Der Pflanzen Band₂, p. 271, Verlag Basel and Stuttgart, 1963.
6. George Watt, A Dictionary of the Economic Products of India Vol. 1, p. 343, Printed by the Superintendent of Govt. Printing, India 1889.
7. R.N. Chopra, S.L. Nayar, I.C. Chopra, Glossary of Indian Medicinal Plants, p. 28, Council of Scientific and Industrial Research, New Delhi, 1956.
8. R. Kirtikar and B.D. Basu, Indian Medicinal Plants Vol.2, p. 1287, Indian Press, Bahadurganj, Allahabad, (1918).

9. P.S. Rao, B.M. Beri and R.P. Budhiraja, J. SCI. IND. RES. (India) 10B, 261 (1951).
10. P.S. Rao and R.P. Budhiraja, J. SCI. IND. RES. 11B, 209 (1952).
11. P.S. Rao and K.G. Gakhar, Proc. Indian Acad. Sci. 36A, 70 (1952).
12. (i) S.B. Rao, Indian J. Pharm. 12(2), 51 (1950).
(ii) S.B. Rao, ibid. 14, 131 (1952).
13. S.S. Subramanian and A.G.R. Nair, Curr. Sci. 37(10), 287 (1968).
14. A.B. Ladge and J.L. Bose, Indian J. Chem. 8, 588 (1970).
15. S.S. Subramanian and A.G.R. Nair, Curr. Sci. 38(17), 414 (1969).
16. The Wealth of India - A Dictionary of Indian Raw Materials and Industrial Products, p. 131, Council of Scientific and Industrial Research, New Delhi, 1948.
17. K.R. Kirtikar and B.D. Basu, Indian Medicinal Plants Vol. IV, p. 2502, Reprint edition, edited, revised, enlarged and mostly rewritten by E. Elatter, J.F. Caius and K.S. Mhaskar, 1975.

18. J.L. Wichers and B. Tollens, J. Landw. 58, 101;
C.A. 4i 3081².
19. J.L. Wichers and B. Tollens, J. Landw. 58, 113;
C.A. 4i 3081⁴.
20. N. Haensel, Geschäftsbericht, April-Sept. through
Chem. Zeutr. 11, 1556 (1909).
21. P.S. Rao and O.N. Rozdon, Proc. Indian Acad. Sci.
31 A, 441 (1950).
22. L.N. Vanquelin and P.J. Robiquot, Ann. Chim. 57, 88
(1806).
23. R. Goldberg, Phytochemistry 8, 1783 (1967).
24. N.E. Hehner, Chem. News 116, 296 (1917).
25. W.E. Cake and H.H. Bartlett, J. Boil. Chem. 51, 93
(1922).
26. A. Winterstein and U. Ehrenberg, Z. Physiol. Chem.
207, 25 (1932).
27. G. Tanret, Compt. rend. 149, 48; C.A. 3 : 2677⁴.
28. B. Tollens, J. Landw. 59, 429; C.A. 6 : 658².
29. L.E. Hamstad, Medd. Norsk. Farm. Selsk. 4, 52 (1942).

30. E.F. Jansen, J. Biol. Chem. 176, 657 (1948).
31. E.W. Juries, Z. Levenson, Uhtersuch Forsch. 132, 193 (1966).
32. U.U. Okonenko and P.E. Krivenchuk, Farm. Zn.(kiev) 21, 44 (1966).
33. F.J. Francis, J. Food Sci. 32, 430 (1967).
34. N. Shiomi, J. Yamada, Agric. Biol. Chem. 40, 567 (1976).
35. G.M. Goryanu, V.V. Krokhmalyuk and P.K. Kintya, Khim. Prir. Soedin 3, 400 (1976); C.A. 85 : 90178w.
36. G.V. Luzure'vskii, G.M. Goryanu and P.K. Kintya, Dokl. Akad. Nauk. SSSR. 231, 1479 (1976), C.A. 86 : 68399t.
37. Rodd's Chemistry of Carbon Compounds. II E, p. 18, second edition, 197. Elsevier Publishing Co., Amsterdam, Netherlands.
38. R. Tschesche, G. Ludke and G. Wulff, Tetrahedran Lett. 29, 2785 (1967).
39. S. Sakamura, Y. Obata, I. Nizuma, K. Nakamura and S. Watanabe, Nippon Shokuhin Kogyo Gakkaishi 14, 491 (1967).

40. G.Y. Held, D. Vagujfalvi and F. Uresch, Phytochemistry 8, 493 (1969).
41. J.J. Doesberg and A. Meijen, Voeding 25, 258 (1964).
42. E.G. Gleim, D.K. Tressler and F. Fenton, Food Research 9, 471 (1944).
43. C. Dame Jr., C.O. Chichester and G.L. Marsh, Food Research 22, 673 (1957); ibid. 24, 20 (1959).
44. W. Schuphan, Qualitas Plant et Materiae Vegetabiles 6, 199 (1960).
45. Jackirmow - Barras Norelly, Phytochemistry 1261, 1331 (1973); C.A. 79 : 50759 f.
46. O.S. Giordano and A. Gonzalez, An. Real Soc. Espan. Fis. Quim. Ser B 63, 945 (1967).
47. F. Elze, Chem. Ztg. 41, 842 (1917).
48. K.R. Kirtikar and B.D. Basu, Indian Medicinal Plants Vol.IV, p. 2499, Reprint edition 1975.
49. P.S. Rao and O.S. Rozdon, Proc. Indian Acad. Sci. 31A, 441 (1950).
50. P.S. Rao, O.S. Rozdon and R.P. Budhiraja, Proc. Indian Acad. Sci. 32A, 264 (1950).

51. V.V. Palasi, Farmacognosia (Madrid) 8, 305 (1948).
52. J.F.L. Candela and V.V. Palasi, Anales real soc. espan. fis. y. quim. 47B, 309 (1951).
53. C.A. Davila and F.M. Panizo, Can. J. Chem. 38, 388 (1960).
54. P.S. Rao, R.M. Beri and R.P. Budhiraja, J. SCI. IND. RES. 11B, 127 (1952).
55. P.S. Rao and K.L. Gakhar, Proc. Indian Acad. Sci. 35A, 310 (1950).
56. S.C. Sharma, R. Chand and O.P. Sati, Pharmazie 35(11), 711 (1980).
57. (i) S.C. Sharma, R. Chand and O.P. Sati, Phytochemistry 21(8), 2075 (1982)
(ii) S.C. Sharma, R. Chand, Bhatti, O.P. Sati, Planta Medi. 46(1), 48 (1982).
58. R.F. Diaz, R.F. Berriere and A. Gonzalez, An. Real Soc. Espan. Fis. Quim. Ser B 63, 927 (1967).
59. S.C. Sharma, R. Chand and O.P. Sati, Curr. Sci. 51, 280 (1982).

60. S.C. Sharma, R. Chand and O.P. Sati, Phytochemistry 21(7), 1711 (1982).
61. Masashi Tomoda and Noriko Satoh, Chem. Pharm. Bull. 22(10), 2306 (1974).
62. T. Kobayashi, T. Tomimori, T. Nakajima and N. Yohogi, Yakugaku Kinkyu 30, 477 (1958).
63. T. Kawasaki, T. Komori, T. Nohara, I. Hosokawa and K. Mahashi, Chem. Pharm. Bull. 22, 2164 (1974).
64. T. Okanishi, A. Akahari, F. Yasuda, T. Yasuyoshi, T. Iwao, Chem. Pharm. Bull 23(3), 575 (1975).
65. S. Kiyosawa, M. Hutoh, T. Komari, T. Nohara, I. Hosokawa, and T. Kawasaki, Chem. Pharm. Bull. 16, 1162 (1968).
66. Tenji Konishi and Junzo Shoji, Chem. Pharm. Bull. 27(12), 3086 (1979).
67. M. Tomoda, N. Satoh, M. Tanaka, Y. Kyoritsu, Daigaku Kenkyu Nempo 20, 9 (1975).
68. Bhagwan Dash and Lalitesh Kashyap, Materia Medica of Ayurveda based on Ayurveda Saukhyam of Tadarananda, p.469, Concept Publishing Co., New Delhi, 1980.

69. Medicinal Plants of India Part I, p.102, Indian Council of Medicinal Research, New Delhi, 1976.
70. Astangahridaya, A.M. Kunte, 6th Edn. Pandurang Jawaji, Nirnaya Sagar Press, Bombay (1939).
71. The Charaka Samhita, J.T. Acharya, 3rd Edn., Satyabhamabai Pandurang, Nirnaya Sagar Press, Bombay (1941).
72. Susrut Samhita, S.G. Ghanekar, Motial Banaresidas, Varanasi (1960).
73. Dr.K.M. Nadkarni, Indian Mat. Med. Vol.II. p.154.
74. R.N. Chopra, Indigenous Drugs of India, p.566.
75. R.N. Khorry, Materia Medica of India and their therapeutics, Vol.II.
76. Sushrut Samhita Sutrasthan, Chap. 26.
77. Bhagwan Dash and Lalitesh Kashyap, Diagnosis and Treatment of Diseases in Ayurveda based on Ayurveda Saukhyam of Todarananda Part 2, p.321, Concept Publishing Co., New Delhi, 1982.
78. U.K. Kanitkar, P.S. Dange and G.S. Pendse, J. Res. Indian Med. 3, 123 (1969).

79. U.K. Kanitkar, P.S. Dange and G.S. Pendse, J. Res. Indian Med. 3, 138 (1969).
80. B.B. Gaitonde, P.B. Sabnis and M.H. Jetmalani, Indian J. Exptl. Biol. 6, 55 (1968).
81. M.H. Jetmalani, P.B. Sabnis and B.B. Gaitonde, J. Res. Indian Med. 2, 1 (1967).
82. B.B. Gaitonde and M.H. Jetmalani, Arch Int. Pharmacodyn 179, 121 (1969).
83. B.B. Gaitonde and M.H. Jetmalani, Proc. II Indo Soviet Sym. Chem. Nat Products including Pharmacology, p.148, New Delhi, 1969.
84. B.B. Gaitonde, Realms of Ayurveda, Pandit Shiv Sharma, p.248, New Delhi, 1979.
85. R.N. Roy, S.R. Chavan, S. Bhagwager, N.K. Datta and N.S. Iyer, Indian J. Pharm. 30, 289 (1968).
86. R.N. Roy, S. Bhagwager, S.R. Chavan and N.K. Datta, J. Res. Indian Med. 6, 132 (1971).
87. M.L. Dhar, M.M. Dhar, B.N. Dhawan, B.N. Mehrotra and C. Roy, Indian J. Exptl. Biol. 6, 232 (1968).
88. P.S. Dange, U.K. Kanitkar and G.S. Pendse, Planta Medica 17, 393 (1969).

S U M M A R Y

The chemistry of genus Asparagus is reviewed. These species contain carbohydrates, saponins, sapogenins, amino acids, colouring matter, inorganic materials etc. Sarsasapogenin is the most commonly occurring aglycone.

Also background work on Asparagus racemosus Willd. is described. The pharmacognostic identification, various extracts of shatavari and their pharmacological screening are described.