

## 6.1. In Vivo Efficacy Study

### 6.1.1 Description of the Methods

This study was performed in 8-10 week old female SCID mice. Mice were housed in individually ventilated cages and given pelleted food (Teklad Lab Animal Diets, Harlan Laboratories, USA) and water ad libitum in a temperature ( $25\pm 3^{\circ}\text{C}$ ) and humidity (50–70%) controlled environment with a 12-h/12-h dark-light cycle. All animal experimentations were carried out in accordance with CPCSEA (Committee for the Purpose of Control and Supervision of Experiments on Animals) guidelines, using (IAEC) Institutional Animal Ethics Committee approved protocols in an AAALAC International (Association for Assessment and Accreditation of Laboratory Animal Care International) accredited facility which complies with the National Institutes of Health guidelines for care and use of animals.(1–3)

SCID mice were injected subcutaneously on the right flank with cultured A549 ( $5 \times 10^6$  cells in 200  $\mu\text{L}$  of RPMI 1640 with 33% Matrigel [BD Biosciences, San Jose, CA]. When xenograft tumor volumes, reached 150  $\text{mm}^3$  (group mean), mice were divided into four groups (n=6) and received the following agents either alone or in combination as specified by the experimental protocols: vehicle or Taxotere® (20mg/kg), docetaxel nano particle (20mg/kg) or cetuximab conjugated docetaxel nanoparticle once weekly (QW) via lateral tail vein. Tumor dimensions were assessed twice weekly using an electronic digital caliper. Tumor volume was calculated as ( $\text{width}^2 \times \text{length}/2$ ). Body weight was recorded twice weekly as an index of toxicity.(4,4,5)

### 6.3. Results and Discussion

To demonstrate the antitumor efficacy, Taxotere®, Docetaxel NPs and Cetuximab Conjugated Docetaxel NPs were injected in human lung cancer-bearing mice via lateral tail vein. In animals bearing established A549 tumors, docetaxel (Taxotere® 20 mg/kg, QW), a standard-of-care chemotherapy agent in NSCLC, significantly inhibited tumor growth compared with vehicle.(6–9) Treatment with Taxotere caused 60.3% inhibition in tumor growth on day 21 compared to vehicle control. Similar to Taxotere similar dosing regimen of docetaxel nano particles showed significant reduction in tumor growth compared to vehicle control. When mice were injected with 20 mg/Kg Docetaxel NPs, demonstrated an enhanced antitumor efficacy compared to the Docetaxel alone. The anti-tumor effect of docetaxel nanoparticle was associated with the body

weight loss may be due to the availability or tissue penetration in the different tissues. However, when combined at the same dose with cetuximab exerts significantly greater antitumor effect than that observed with single agent alone.(4)(4,10,11) Treatment with docetaxel NPs caused 72.5% reduction on tumor volume while 81% inhibition was observed in cetuximab-conjugated docetaxel NPs. Surprisingly anti-tumor effect was associated with lack of body weight loss in the cetuximab-conjugated docetaxel NPs indicating that the drug was targeted to site of action. Thus, Nano-sized docetaxel can provide advantages of reducing the high dose dependent toxicity of anticancer drugs while, at the same time increasing their anticancer efficacy.(4,12,13) Furthermore, this result clearly indicate that antibody (Cetuximab) conjugation to Docetaxel NPs is successful in inhibiting tumor growth while lower toxicity effect.

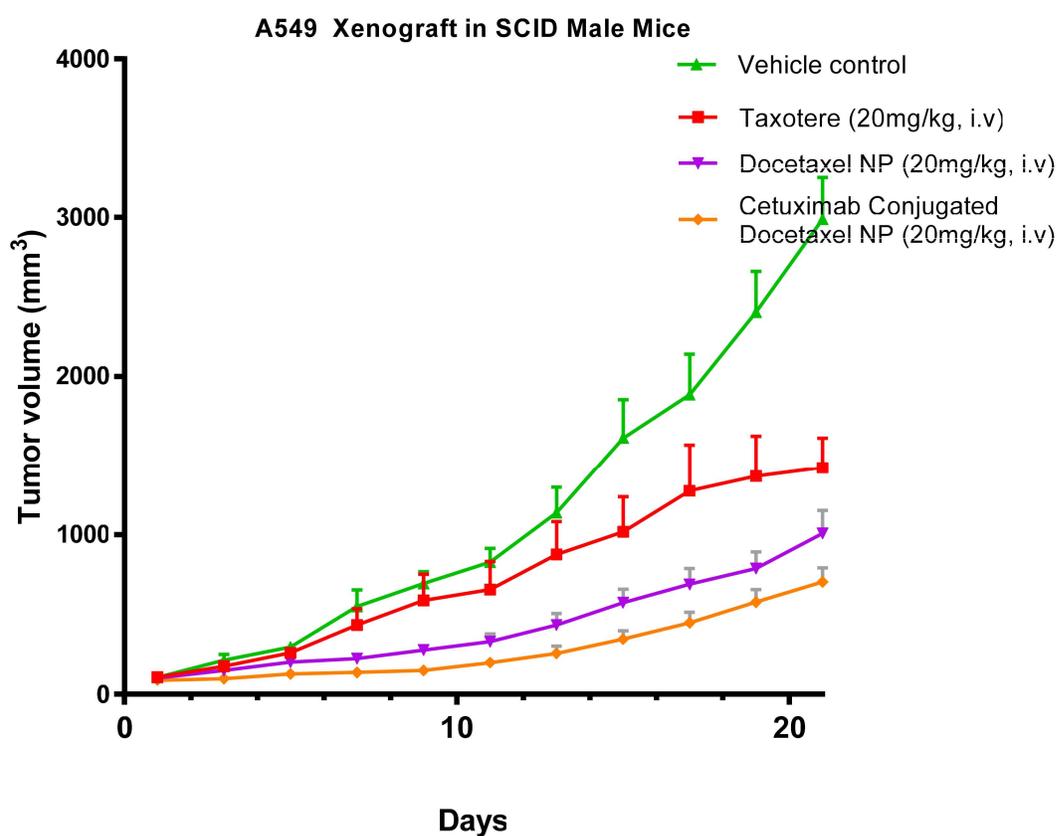
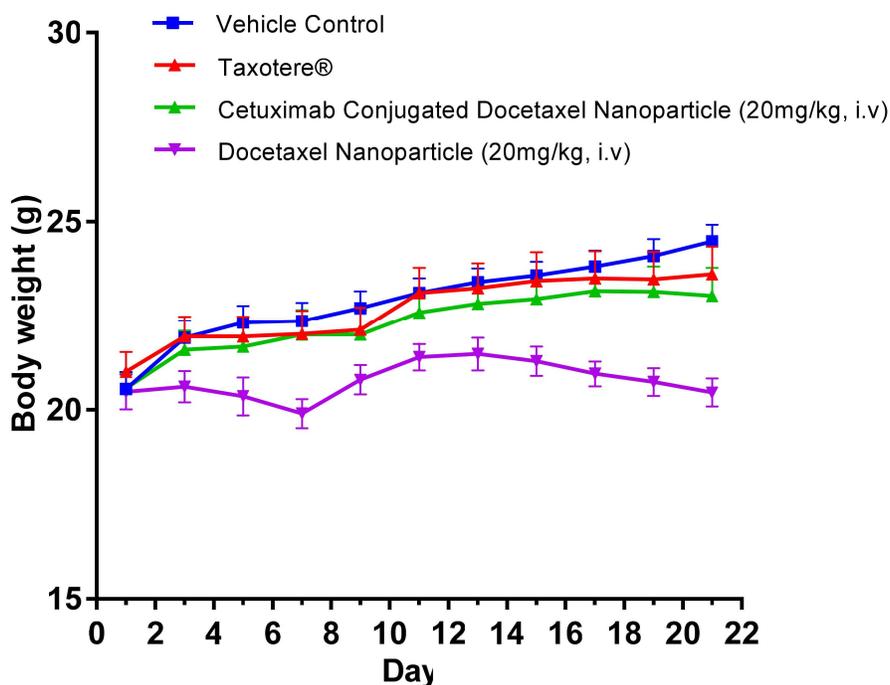


Figure 6.1

**A549 Xenograft model: Tumor volume vs Days**

**A549 Xenograft in SCID Male Mice****Figure 6.2 A549 Xenograft model: Body weight vs Days****Conclusions**

Our data show that docetaxel nanoparticles exert a strong antitumor activity over marketed docetaxel in A549 NSCLC subcutaneous xenograft model. When conjugated with cetuximab with docetaxel nanoparticle was significantly greater than docetaxel alone treatment in the xenograft models. Toxicity was significantly reduced in docetaxel nanoparticles conjugated with cetuximab compared to docetaxel nanoparticle alone.

**Reference List**

1. Chen Y, Liu G, Guo L, Wang H, Fu Y, Luo Y. Enhancement of tumor uptake and therapeutic efficacy of EGFR-targeted antibody cetuximab and antibody-drug conjugates by cholesterol sequestration. *Int J Cancer*. 2015;136(1):182–94.
2. Jain A, Thakur K, Kush P, Jain UK. Docetaxel loaded chitosan nanoparticles: formulation, characterization and cytotoxicity studies. *Int J Biol Macromol* [Internet]. 2014 Aug [cited 2016 Feb 15];69:546–53. Available from: <http://www.sciencedirect.com/science/article/pii/S0141813014004127>
3. Kundu AK, Chandra PK, Hazari S, Pramari Y V, Dash S, Mandal TK. Development and optimization of nanosomal formulations for siRNA delivery to the liver. *Eur J Pharm Biopharm Off J Arbeitsgemeinschaft für Pharm Verfahrenstechnik eV* [Internet]. 2012 Feb [cited 2016 Feb 25];80(2):257–67. Available from: <http://www.sciencedirect.com/science/article/pii/S0939641111003213>
4. Kurai J, Chikumi H, Hashimoto K, Yamaguchi K, Yamasaki A, Sako T, et al. Antibody-dependent cellular cytotoxicity mediated by cetuximab against lung cancer cell lines. *Clin Cancer Res* [Internet]. 2007;13(5):1552–61. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/17332301>
5. Cheng J, Teply BA, Sherifi I, Sung J, Luther G, Gu FX, et al. Formulation of functionalized PLGA – PEG nanoparticles for in vivo targeted drug delivery. *Biomaterials*. 2007;28:869–76.
6. Bissery MC, Gu??nard D, Gu??ritte-Voegelein F, Lavelle F. Experimental antitumor activity of taxotere (RP 56976, NSC 628503), a taxol analogue. *Cancer Res*. 1991;51(18):4845–52.
7. Clarke SJ, Rivory LP. Clinical pharmacokinetics of docetaxel. *Clin Pharmacokinet* [Internet]. 1999;36(2):99–114. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/10092957>
8. Zhao P, Astruc D. Docetaxel Nanotechnology in Anticancer Therapy. *ChemMedChem*.

2012. p. 952–72.
9. Ciccolini J, Catalin J, Blachon M., Durand A. Rapid high-performance liquid chromatographic determination of docetaxel (Taxotere) in plasma using liquid–liquid extraction. *J Chromatogr B Biomed Sci Appl* [Internet]. 2001 Aug [cited 2016 Feb 21];759(2):299–306. Available from: <http://www.sciencedirect.com/science/article/pii/S0378434701002389>
  10. Kutty RV, Feng S-S. Cetuximab conjugated vitamin E TPGS micelles for targeted delivery of docetaxel for treatment of triple negative breast cancers. *Biomaterials* [Internet]. 2013 Dec [cited 2016 Feb 15];34(38):10160–71. Available from: <http://www.sciencedirect.com/science/article/pii/S0142961213011265>
  11. Yi C, Ruan C, Wang H, Xu X, Zhao Y, Fang M, et al. Function characterization of a glyco-engineered anti-EGFR monoclonal antibody cetuximab in vitro. *Acta Pharmacol Sin* [Internet]. 2014;35(11):1439–46. Available from: <http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=4220073&tool=pmcentrez&rendertype=abstract>
  12. Maya S, Sarmiento B, Lakshmanan V-K, Menon D, Seabra V, Jayakumar R. Chitosan cross-linked docetaxel loaded EGF receptor targeted nanoparticles for lung cancer cells. *Int J Biol Macromol* [Internet]. 2014 Aug [cited 2016 Feb 13];69:532–41. Available from: <http://www.sciencedirect.com/science/article/pii/S0141813014003924>
  13. Perri F, Muto P, Schiavone C, Ionna F, Aversa C, Maiolino P, et al. 583 Induction Chemotherapy with Docetaxel, Cisplatin and Capecitabine, Followed by Combined Cetuximab and Radiotherapy in Patients with Locally Advanced Inoperable Head and Neck Cancer. Preliminary Results of a Phase I/II Study. *Eur J Cancer* [Internet]. 2012 Nov [cited 2016 Feb 11];48:178–9. Available from: <http://www.sciencedirect.com/science/article/pii/S0959804912723808>